COMPARATIVE STUDY OF ESSENTIAL OIL COMPOSITION OF FRESH AND DRY PEEL AND SEED OF CITRUS SINENSIS (L) OSBECK VAR SHAMUTI AND CITRUS PARADISI MACFADYEN VAR MARSH

Olonisakin Adebisi

Department of Chemical Sciences
Adekunle Ajasin University, PMB 001. Akungba-Akoko. Ondo-State. Nigeria.

E-mail: adeoloni@yahoo.com
(Received: 2nd June, 2014; Accepted:17th July, 2014)

ABSTRACT

Citrus essential oils have an impressive range of food and medicinal uses. In this study investigation has been conducted on the variation in the yield, chemical composition and their identities in oils isolated from fresh and air-dried peel and seed of orange (*Citrus sinensis*) and grape(*Citrus paradisi*) planted in a cocoa farm. The yield of solvent-extracted essential oils from the fresh peel and seed ranged between 0.31 and 1.01%, while the yield in the air-dried peel and seed of the two different citrus samples ranged between 0.98 and 2.30%. The four major compounds present in all the oils are limonene, myrcene, alpha terpinene and camphene which ranged between 74.97 - 90.58%, 5.19 - 10.41%, 0.14 - 4.00% and 0.05 - 3.87%, respectively in fresh peel and seed. In the air-dried peel and seed their values ranged between 58.64 - 77.30%, 0.08 - 5.04%, 0.05 - 3.68% and 0.02-4.88%, respectively for the four compounds. The fresh peel and seed have lower yield but contain higher percentage concentrations of major compounds that serve as compound identification for the citrus family. Air-dried peel and seed have alcohol components like spathulenol (9.78 - 15.13%), linalool (5.05 - 9.27%), nerol (7.98 - 8.60), alpha terpeniol (1.06 - 1.15%) and farnesol (1.54 - 1.66%) which were not present in fresh samples of the two different citrus samples. Apart from concentration differences, the results of this study are similar to other research work in other regions of the world on citrus; the only difference was the identification of camphene in this study but which was not found in citrus located in other regions of the world under consideration.

Keywords: Citrus peel, Seed, Fresh, Air-dried, Essential Oil.

INTRODUCTION

Citrus essential oils are obtained from various Citrus species found within the family Rutaceae and the subfamily Aurantioideae; these include about 17 species distributed throughout the tropical and temperate regions (Davies and Albrigo, 1994; Shaw, 1977). They are evergreen trees that give fruits of different forms and sizes (from round to oblong), which are full of fragrances, flavor and juice. The citrus genus includes various species; oranges (Citrus sinensis), limes (Citrus aurantifolia), tangerine (Citrus reticulate), lemons (Citrus limon) and grapefruit (Citrus paradisi) (Burcu et al., 2011; Mohammed et al., 2010). Citrus is one of the most important commercially-grown crops in the world. The world production of citrus was estimated to be 51.8 million metric tonnes during 2013 with Brazil, China and United States as the first three leading producers in the world and Egypt, South Africa and Morocco, respectively as the leading producers in the African continent (USDA, 2014). Citrus essential oil are a mixture of volatile compounds and mainly consists of monoterpene hydrocarbons of about 70-95% along with smaller

amount of sesquiterpene hydrocarbons which are responsible for a characteristic flavor (Muhammad, 2006).

Although, the fruits are mainly used for dessert, it has significant economic value for its essential oil (EO) due to their aromatic compounds (Minh et al., 2002). Citrus flavours are used in beverage, confectionary, cookies and desserts (Buchel, 1989; Dharmawan et al., 2007). The exocarps of C. reticulata and C. sinensis are used for flavorings of liquor. The composition of the oil is significantly affected by the ripeness of fruits, vegetative stage of plant, storage condition and extraction method (Njoroge et al., 2006; Venkateshwarlu and Selvaraj, 2000). The quality and the odour of the oil are influenced by the limonene content which may vary in the different agro-climatic conditions (Dharmawan et al., 2007).

Like many other plant families, Rutaceae has been reported to be a source of potent botanical insecticide. Peel and seed solvent extract of citrus plant have shown insecticidal activity against several coleopteran and dipteran species (Su *et al.*,

1972; Greany et al., 1983; Sheppard 1984; Salvatore et al., 2004). Previously it has been established that *C. aurantium* peels contain secondary metabolites with insecticidal activity against *B. oleae* adult (Siskos et al., 2007).

Citrus essential oils have been applied in many products, such as foods, beverages, cosmetics and medicines, as flavouring agents as well as for aromatherapy. They are also used for their germicidal, antioxidant and anticarcinogenic properties (Guenther, 1948; Mukhopadhyay, 2000). The active constituents exist in citrus EOs, such as limonene, α -pinene, β -pinene and α -terpinolene which exhibit a wide spectrum of antimicrobial activity, as reported by many studies in other plants (Jirovetz *et al.*, 2005; Magwa *et al.*, 2006; Skocibusic *et al.*, 2006; Matasyoh *et al.*, 2007).

The biological properties of EOs extracted from medicinal plants may show differences depending upon the distinctness in cultivation, origin, vegetative stage and growing seasons of the plants and geographical location (Deans *et al.*, 1992; Muller-Riebau *et al.*, 1995; Kustrak *et al.*, 1996; Leung and Foster, 1996; Milos *et al.*, 2000; Jerkovic *et al.*, 2001). In addition, other factors that can affect chemical composition are extraction process, stage of harvesting and post-harvest processing (Dean *et al.*, 1992).

In this study, the main aim was to obtain essential oils from orange (*C. sinensis*) and grapefruit (*Citrus paradisi* L.) peel and seed using solvent extraction method under fresh and air dried conditions in order to compare the yield and chemical composition. Also this study will obtain information about the chemical variability of the peel and seed from this region of Nigeria and compare to result from other regions.

MATERIALS AND METHODS Sample Collection and Preparation

The fresh fully matured ripened fruits of two species of Citrus: Sweet orange (*C. sinensis*) (CS) and grape (*C. paradisi*) (CP) were collected from a cocoa farm in Isinbode-Ekiti, Ekiti-State. Nigeria. The citrus varieties were identified in the Department of Plant Science and Biotechnology of Adekunle Ajasin University, Akungba Akoko. Ondo State, Nigeria. The two citrus fruit species

were washed free of sand over tap water. The fruits were then peeled off carefully with the help of a sharp razor blade to avoid any damage of oil glands. Due to practical reasons, the *Citrus* peels under testing were processed under two categories: one portion used as fresh, the other airdried at ambient temperature (30 °C) were ground and extraction took place immediately while the other portion was air-dried.

Extraction

Precisely, 200 g of each of the samples was packed into a Soxhlet apparatus and extracted exhaustively with 750 ml of Hexane for 3 h. The solvent was recovered using a rotary evaporator. The oils were made to be moisture free by filtering through the anhydrous sodium sulphate. The oils were transferred into brown sample bottles and stored in the refrigerator until ready for analysis

Analysis of the Components

The volatile constituents were identified by gas chromatography technique using direct injection in the split mode with a split ratio of 20:1 under the following conditions: Hewlet-Packard 6890 equipped with a quartz capillary column; 30 mm x 0.25 mm internal diameter and 0.25 µm film thickness was used. Hydrogen was the carrier gas at 1.0 ml/min flow rate; oven temperature, 40°C to 200°C at a rate of 5°C/min then held isothermal for 2 min. Injector temperature and volume are 150°C and 1.0 μl, respectively. The ionization of the sample components was performed on E.I mode (70eV). The identification of different constituents was performed by comparison of their retention time and mass spectra with those of the library.

RESULTS AND DISCUSSION

The yield and colour of the essential oil from the peel and seed samples are presented in Table 1. The yield ranged between 0.31-1.01% in fresh peel and seed with peel having the lowest values. The air-dried peel and seed had the yield ranged between 0.98 - 2.30%. The fresh samples generally had low yield when compared to dry samples, this may be due to the high water content of the materials. The seed of both citrus had higher yield when in dry condition than in fresh. According to Williams, 1996, depending on the variety of the fruit peel or seed, citrus yields essential oil content

of between 0.5 and 5.0%.

Table 1: Essential Oil Yield and Colour of Essential Oil from Peel and Seed of CS and CP

Species	Condition	Yield %v/w	Colour		
C. sinensis (peel)	Fresh	0.40	Light green		
C. sinensis (seed)	Fresh	0.67	Light yellow		
C.paradisi (peel)	Fresh	0.31	Light green		
C.paradisi (seed)	Fresh	1.01	Light yellow		
C. sinensis (peel)	Dry	1.09	Dark green		
C. sinensis (seed)	Dry	1.73	Dark yellow		
C.paradisi (peel)	Dry	0.98	Dark green		
C.paradisi (seed)	Dry	2.30	Dark yellow		

The yield obtained from this study with respect to the effects of drying condition on peel and seed essential oil are in agreement with the findings of Asekun et al. (2007) who also investigated higher oil contents from oven-dried samples of Helichrysum odoratissimum in comparison with fresh material. Asekun et al. (2006) had earlier studied the effects of drying on the yield and chemical composition of essential oil from the aerial parts of Leonotis leonurus and found that the oils derived from sun-dried plant material had better yield than those from the air and oven-dried materials. Some other reports in the literature also revealed considerable effects of drying on the yield and characteristics of the essential oils (Rahula et al., 1973; Laranja et al., 2003). Furthermore, the yield obtained in this work which ranged between 0.31 -2.30% is slightly higher than the one obtained by Tu et al. (2002) (0.20 -2.0%) and Kamal et al. (2001) (0.24 - 1.07%). This may be due to the method of extraction in their studies, steam distillation was used and solvent extraction was used in this study. In essential oil extraction methods, solvent extraction technique has been found to have higher yield than other methods.

The results of the essential oil composition of the fresh and air-dried peel and seed of C. sinensis and C. paradisi are shown in Table 2. Peel and seed oils consisted almost exclusively of hydrocarbons, with limonene as major component. Limonene, myrcene, α-pinene and camphene were the main components in the EO. Linalool is the most abundant oxygenated monoterpene. The fresh peel and seed were found to contain high

concentration of limonene in all the samples when compared to air-dried samples, for instance limonene content in the fresh peel ranged between 88.32 to 90.58% in CP and CS, respectively, while in fresh seed limonene content ranged between 74.97 – 81.73% for CP and CS, respectively. Limonene percentage concentration ranged between 58.64 - 65.95%, respectively for air-dry peel of CP and CS, while it ranged between 67.92 to 77.30% for CP and CS, respectively for air-dried seed. Myrcene is another compound that is present in all the samples and it ranged between 0.08% minimum in C. sinensis dry peel (CSDP) to maximum of 10.41% in C. paradise fresh seed (CPFS). Camphene ranged between 0.02 to 4.88%, respectively in CSDP and CPDS. The fresh peel sample is however higher in limonene content than the fresh seed. Moreover the fresh peel of CS was found to contain more number of terpenoids than CP fresh peel, for instance α -thujene, γ terpinene, citral and neral were present in C. sinensis fresh peel (CSFP) but not in C. paradise fresh peel (CPFP) and the fresh seed of the two citrus almost have the same number for terpenoids except linalool that is present in CSFS. In all the samples, it was observed that air-dried samples of CS and CP peel have the highest number of compounds. Compounds like nerol, α terpineol, spathulenol and farnesol (alcohol) were present in these two oils at higher concentration in C. paradise dry peel (CPDP) than CSDP but were absent in all the remaining oils. This alcohol content ranged between 28.91% to 35.81% in CS and CP, respectively. The general result indicates that fresh sample has more main components in

higher concentration than the dry one. This may be due to the volatile nature of terpenoids during the course of its drying. The alcohol content that was not found in the fresh samples was seen in the dry sample of the citrus most especially the dry peel.

Table 2: Essential Oil Composition (%) of Fresh and Air-dry Peel and Seed of *C. sinensis* and *C. paradisi*.

Compound	CSFP	CPFP	CSFS	CPFS	CSDP	CPDP	CSDS	CPDS
-	%	%	%	%	%	%	%	%
Cymene	-	-	_		2.09	2.26	1.82	1.37
α-phellandrene	-	-	0.27	0.04	-	-	0.03	0.72
α- terpinene	0.14	1.86	4.00	6.62	0.05	0.06	1.59	3.68
β-ocimene	-	-	0.27	0.02	-	-	0.03	0.02
Camphene	0.17	0.05	3.87	3.74	0.02	0.03	1.38	4.88
Terpinolene	-	-	0.22	0.02	-	-	0.03	0.02
Sabinine	-	-	0.36	0.03	-	-	0.04	0.02
Limonene	90.58	88.32	81.73	74.97	65.95	58.64	77.30	67.92
α-pinene	2.27	2.08	0.93	3.33	-	1.36	3.17	2.31
β-pinene	-	0.01	0.58	0.05	-	-	0.08	0.05
Myrcene	5.19	7.00	5.35	10.41	0.08	0.86	2.65	5.04
Thujene	0.10	-	0.21	0.01	0.14	0.16	6.38	6.84
γ-terpinene	0.02	-	0.96	0.90	-	-	0.21	0.67
Citral	0.03	-	-	-	0.03	0.03	-	-
Geranial (nera)	0.49	-	-	-	0.57	0.62	-	-
Linalool	-	-	0.04	-	8.57	9.27	5.05	6.78
Nerol	-	-	-	-	7.96	8.60	-	-
α-terpineol	-	-	-	-	1.06	1.15	-	-
Spathulenol	-	-	_	-	9.97	15.13	-	-
Farnesol	-	-	_	-	1.54	1.66	-	-

CSFP = C. sinensis fresh peel, CPFP = C. paradisi fresh peel, CSFS = C. sinensis fresh seed, CPFS = C. paradisi fresh seed, CSDP = C. sinensis dry peel, CPDP = C. paradisi dry peel, CSDS = C. sinensis dry seed, CPDS = C. paradisi dry seed.

Table 3: Essential Oil	Composition of	Fresh Peel and	Seed of Present Str	udy and Fresh Peel of	Other Region

Compounds	CSFP	CSFS	CPFP	CPFS	ITCS	FLCS	TUCS	PKCP1	IRCS	PKCP2	GRCS
	%	%	%	%	%	%	%	%	%	%	%
α-terpinene	0.14	4.00	1.86	6.62	-	-	1.0	2.11	0.20	-	-
Camphene	0.17	3.87	0.05	3.74	-	-	-	-	-	-	-
Sabinene	-	0.36	-	0.03	1.00	0.24	0.06	-	0.5	-	0.2
Limonene	90.50	81.73	88.32	74.97	93.67	95.17	88.60	86.27	90.50	76.28	94.20
α-Pinene	2.27	0.93	2.08	3.33	0.67	0.42	0.70	1.26	1.40	1.26	0.80
β-Pinene	-	0.58	0.01	0.05	-	-	1.20	-	-	5.45	-
Myrcene	5.19	5.35	7.00	10.41	2.09	1.86	0.90	6.28	2.50	1.30	2.80
Thujane	0.10	0.21	-	0.01	-	-	-	0.15	-	-	-
Octanal	-	-	-	-	-	-	-	-	-	0.80	-
linalool	-	0.04	-	-	0.31	0.25	0.70	0.02	0.80	2.32	0.80
3-carene	-	-	-	-	0.31	-	0.50	-	-	-	-
Decanal.	-	-	-	-	0.27	o.28	-	-	-	-	

CSFP = C. sinensis fresh peel, CSFS = C. sinensis fresh seed, CPFP = C. paradis fresh peel, CPFS = C. paradise fresh seed, ITCS = Italian C. sinensis (Verzera et al., 2004), FLCS = Florida C. sinensis (Vora, 1983), TUCS = Turkey C.sinensis (Burcu, et al., 2011) PKCP1 = Pakistan C. paradisi 1(Muhammad, et al., 2006), IRCS = Iran C. sinensis (Parvizetal, 2006), PKCP2 = Pakistan C. paradis 2 (), GRCS = Greece C.sinensis(Filitsa, et al., 2013)

Table 3 shows the chemical composition of fresh CS and CP peel and seed of this study and that of some fresh peels from other regions of the world. In all the essential oil under consideration, it was noticed that limonene, myrcene, pinene and linalool were present in all the geographical locations in the order of limonene > myrcene > pinene. Limonene percentage value ranged between 74.97- 95.17% in CPFS and Florida sample (Bauer and Surburk, 2001), respectively and this is in agreement with Muhammad et al. (2006) that stated that limonene value of citrus range between 75- 97%. Myrcene in this study is higher in percentage concentration (2.09-10.41%) than the values of other essential oils from other regions which ranged between 0.90-6.20% in Turkey and Pakistan, respectively (Burcu et al., 2011; Muhammad et al., 2006). The alpha pinene in this study is in higher concentration (0.65 - 3.33%) than other regions which ranged between 0.42-1.26% in Florida and Iran, respectively (Burcu et al., 2011; Parviz et al., 2011).

CONCLUSION

The essential oil yields in dry samples are higher than the fresh but the percentage concentration of the major compounds are higher in the fresh than the dry. Limonene, myrcene, α-terpinene, pinene and camphene were all present in the oils under study but when compared with the other region only camphene were absent in other region but present in this study. Linalool was found in other oils from other region but is only on the dry samples that have linalool in this study with higher concentration than other regions.

REFERENCES

Asekun, O. T., Grierson, D. S. and Afolayan, A. J. 2006. Influence of drying methods on the chemical composition and yield of the oil of Leonotis leonurus. Journal of Scientific Research and Development 10: 61-64.

Asekun, O. T., Grierson, D. S. and Afolayan, A. J. 2007 Characterization of essential oils from Helichrysum odoratissimum using different drying methods. Journal of Applied Science 7: 1005 - 1008.

Buchel, J.A. 1989. Flavoring with citrus oil. Perfumer and Flavorist 14: 22–26.

Burcu, U., Fazli, S., Ozgur, A., Birsen, S.O. and Elif, O. K. 2011 Essential oil composition and antibacterial activity of the grapefruit (Citrus paradisi. L) peel essential oils obtained by solvent-free microwave extraction: comparison with hydrodistillation. International Journal of Food Science and Technology. 46, 1455–1461.

Colman, R.L., Lund, E.D. and Moshonas, M.G. 1969. Composition of Orange Essence

- Oil. Journal of Food Science 34 (6): 610-611.
- Davies, F. S. and Albrigo, L. G. 1994. *Citrus*. CAB International, Wallingford, 1pp.
- Deans, S.G., Svoboda, K.P., Gundidza, M. and Brechany, E.Y. 1992. Essential oil profiles of several temperate and tropical aromatic plants: their antimicrobial and antioxidant activities. *Acta Horticulturae* 306: 229–232.
- Dharmawan, J., Kasapis, S., Curran, P. and Johnson, J.R. 2007. Characterization of volatile compounds in selected citrus fruits from Asia. Part I: freshly-squeezed juice. *Flavour and Fragrance Journal* 22: 228–232.
- Filitsa, K., Athanasios, K., Ántonios, M., Dimitrios, p., Moschos, P., Panagiota, P. and Åleanna, T. 2013. Insecticidal activity of plant essential oils against the vine mealybug, *Planococcus ficus. Journal of Insect Science* 13, Article 142.
- Greany, P.D., Styer, S.C., Davis, P.L., Shaw, P.E. and Chambers, D.L. 1983. Biochemical resistance of Citrus to fruit flies. Demonstration and elucidation of resistance to the Caribbean fruit fly. *Anastrepha suspensa*. *Entomol. Exp. Appl.* 34: 40–50.
- Guenther, E. 1948. *The Essential Oils*. Van Nostrand-Reinhold. New York, USA.
- Jerkovic, I., Mastelic, J. and Milos, M. 2001. The impact of both the season of collection and drying on the volatile constituents of *Origanum vulgare* L. ssp. hirtum grown wild in Croatia. *International Journal of Food Science and Technology* 36: 649–654.
- Jirovetz, L., Buchbauer, G., Stoyanova, A.S., Georgiev, E.V. and Damianova, S.T. 2005. Composition, quality control and antimicrobial activity of the essential oil of cumin (*Cuminum cyminum* L.) seeds from Bulgaria that had been stored for up to 36 years. *International Journal of Food Science and Technology* 40, 305–310.
- Kamal, G. M., Anwar, F., Hussain, A. I., Sarri, N. and Ashraf, M. Y. 2011 Yield and chemical composition of Citrus essential oils as affected by drying retreatment of peels *International Food Research Journal* 18(4):1275-1282.
- Kustrak, D., Kuftinec, J., Blazevic, N. and Maffei, M. 1996. Comparison of the essential oil

- composition of two subspecies of *Satureja* montana. Journal of Essential Oil Research 8: 7–13.
- Laranja, D., Mendes, M. F. and Calçada, L. A. 2003.Influência da Secagem na Composição do Óleo Essencial de Citronela, II Simpósio Brasileiro de Óleos Essenciais, IAC, Campinas.
- Leung, A.Y. and Foster, S. 1996. Encyclopaedia of Common Natural Ingredients Used in Foods, Drugs and Cosmetics, 2nd edn. Wiley.New York, USA. 465–466 pp.
- Magwa, M.L., Gundidza, M., Gwerua, N. and Humphrey, G. 2006. Chemical composition and biological activities of essential oil from the leaves of *Sesuvium portulacastrum*. *Journal of Ethnopharmacology* 103:85–89.
- Matasyoh, J.C., Kiplime, J.J., Karubiu, N.M. and Hailstorks, T.P. 2007. Chemical composition and antimicrobial activity of essential oil of Tarchonanthus camphorates. *Food Chemistry* 101: 1183–1187.
- Milos, M., Mastelic, J. and Jerkovic, I. 2000. Chemical composition and antioxidant effect of glycosidically bound volatile compounds from oregano (*Origanum vulgare* L. ssp. hirtum). *Food Chemistry* 71: 79–83.
- Minh Tu, N.T., Thanh, L.X., Une, A., Ukeda, H. and Sawamura, M. 2002. Volatile constituents of Vietnamese pummelo, orange, tangerine and lime peel oils. *Flavour Fragrance Journal* 17: 169–174.
- Mohamed, A. A, El-Emary, G. A. and Ali, H. F. 2010. Influence of some citrus essential oils on cell viability, Glutathione-S-Transferase and Lipid Peroxidation in Ehrlich ascites carcinoma cells. *Journal of American Science* 6 (10):820-826.
- Muhammad, M. A., Salim-ur-rehman, Z. I., Faqir, M.A. and Javaid, I.S. 2006. Genetic variability of essential oil composition in four citrus fruit species. *Pak. J. Botanical* 38 (2): 319-324.
- Mukhopadhyay, M. 2000. Natural Extracts Using Supercritical Carbon Dioxide. CRC Press, New York, USA.
- Muller-Riebau, F., Berge, B. and Yegen, O. 1995. Chemical composition and fungitoxic

- properties to phytopathogenic fungi of essential oils of selected aromatic plants growing wild in Turkey. Journal of Agricultural and Food Chemistry 43: 2262-2266.
- Njoroge, S.M., Mungal, H.N., Koaze, H., Phi, N.T.L., and Sawamura, M. 2006. Volatileconstituents of mandarin Citrus reticulata Blanco peel oil from Burundi. Journal of Essential Oil Research 18: 659–662.
- Parviz, A.A., Mehdi, N., Kambiz, L. and Sakineh, B. 2011. Chemical composition of the essential oils of Citrus sinensis cv. valencia and a quantitative structure-retention relationship study for the prediction of retention indices by multiple linear J. Serb. Chem. Society 76 (12): regression 1627-1637
- Pino, J., Sánchez, M., Sánchez, R. and Roncal, E. 2006. Chemical composition of orange oil concentrates. Nahrung / Food 36 (6): 539-542.
- Rahula, O. B. and Wijesekera, A. L. 1973. Varietal Compositions in the constituents of citronella oil. *Phytochemistry* 12: 2697-2708.
- Salvatore, A., Borkosky, S., Willink, E. and Bardo'n, A. 2004. Toxic effects of lemon peel constituents on Ceratitis capitata. J. Chem. Ecology 30: 323–333.
- Shaw, P. E. 1977. Essential oils. In Citrus Science and Technology. Nagy S., Shaw P. E. and Veldhuis M. K. (Eds.). The AVI Publishing Co. Inc., Westport C.T., 247pp
- Sheppard, D.C. 1984. Toxicity of Citrus peel liquids to the house fly and red imported fine ant. J. Agric. Entomology 1:95–100.
- Siskos, E.P., Konstantopoulou, M.A., Mazomenos, B.E. and Jervis, M. 2007. Insecticidal activity of Citrus aurantium fruit, leaf and shoot extracts against adults of the olive fruit flies. J. Econ. Entomology

- 100 (4):1215 1220
- Skocibusic, M., Bezic, N. and Dunkic, V. 2006. Phytochemical composition and antimicrobial activities of the essential oils from Satureja subspicata Vis. growing in Croatia. Food Chemistry 96: 20-28.
- Su, H.C.F., Speirs, R.D., Mahany, P.G. 1972. Toxicity of Citrus oils to several storedproduct insects: laboratory evaluation. J. Econ. Entomology 65: 1438–1441.
- Tu, M.N.T., Onishi, Y. Choi, H.S., Kondo, Y., Bassore, S.M., Ukeda, H. and Sawamura, M. 2002. Characteristic odor components of Citrus sphaerocarpa Tanaka (Kabosu) cold-pressed peel oil. Journal of Agricultural Food Chemistry 50: 2908-2913.
- United States Department of Agriculture (USDA) (2014). Citrus: world markets trade. Foreign Agriculture Service of Globe Aalysis bulletin.
- Venkateshwarlu, G. and Selvaraj, Y. 2000. Changes in the peel oil composition of Kagzi lime (Citrus aurantifolia Swingle) during ripening. Journal of Essential Oil Research 12: 50 - 52.
- Verzera, A., Trozzi, A., Dugo, G., Di Bella, G. and Cotroneo, A. 2000. Biological lemon and sweet orange essential oil composition. Flavour and Fragrance Journal 19 (6): 544 -548
- Vora, J. D., Matthews, R. F., Crandall, P. G. and Cook, R. 1983. Preparation and Chemical Composition of Orange Concentrates. Journal of Food Science 48 (4): 1197-1199.
- Williams, David G. 1996. The chemistry of essential oils: Micelle Press, Dorset, England